

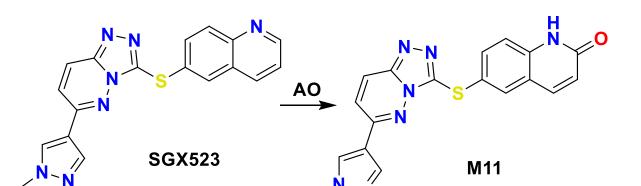
# Liver Humanized Chimeric PXB-Mouse<sup>®</sup> as a Model to Predict Human Renal Toxicity by Aldehyde Oxidase Dependent Metabolism of SGX523

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## Introduction

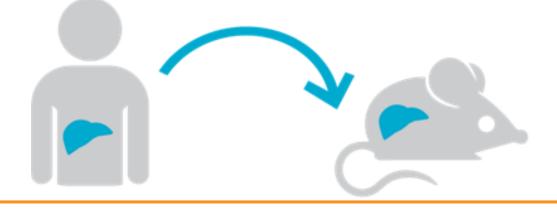
Aldehyde oxidase (AO) is a cytosolic molybdenum-containing hydroxylase that oxidizes a variety of azaheterocycles and has gained the attention of drug discovery programs due to profound species differences in activity and challenges predicting clearance. In addition, metabolites generated by AO are poorly predicted from preclinical studies in rodents and canines, complicating safety assessment programs. SGX523, a selective drug candidate for MET receptor tyrosine kinase, was halted in the clinic due to acute renal failure.<sup>1</sup> The observed clinical findings were unexpected due to SGX523 demonstrating an acceptable safety profile in preclinical toxicology studies in rat and dog. Metabolite profiling of plasma from human patients identified a key AO-mediated circulating metabolite (M11), which was not observed in rats and dogs. Retrospective toxicology studies in monkey (a species with high AO-activity) provided evidence of intratubular crystals of M11 and associated inflammatory changes in kidney tissue, consistent with crystallization of this poorly soluble metabolite in the kidney.<sup>2</sup> As an alternative to utilizing nonhuman primates, the humanized PXB-mouse<sup>®</sup> (Phoenix Bio) has been developed as a predictive model of human metabolism. In effort to assess the suitability of this humanized mouse model for recreating the renal toxicity observed by SGX523, PXB-mice were exposed to SGX523 by single or repeat oral dose daily for up to seven days at 100 mg/kg, followed by analyses of blood chemistry, renal histopathology, and SGX523 and M11 exposure. In addition to measuring effects on serum creatinine (CREAT) and blood urea nitrogen (BUN) as markers of kidney injury, we have included urinary kidney injury molecule-1 (KIM-1) and neutrophil gelatinase-associated lipocalin (NGAL) as more sensitive markers of tubular injury. Upon dosing PXB-mice with SGX-523, elevated levels of serum creatinine, BUN, urinary KIM-1 and NGAL were observed with increasing dose. This correlated with high M11 metabolite/SGX523 ratios in the urine and kidney tissue, in addition to higher circulating exposure of M11. Microscopically, obstructive/retrograde nephropathy was characterized by renal tubular dilatation in a linear distribution and multifocal birefringent amorphous crystals in tubules. This work showcases the utility of KIM-1 and NGAL for early risk assessment of kidney injury for drug candidates, in addition to the utility of a humanized liver mouse model to assess safety of specific human metabolites as an alternative to standard preclinical toxicology species, such as non-human primates.

Figure 1. SGX523 Metabolism by Aldehyde Oxidase

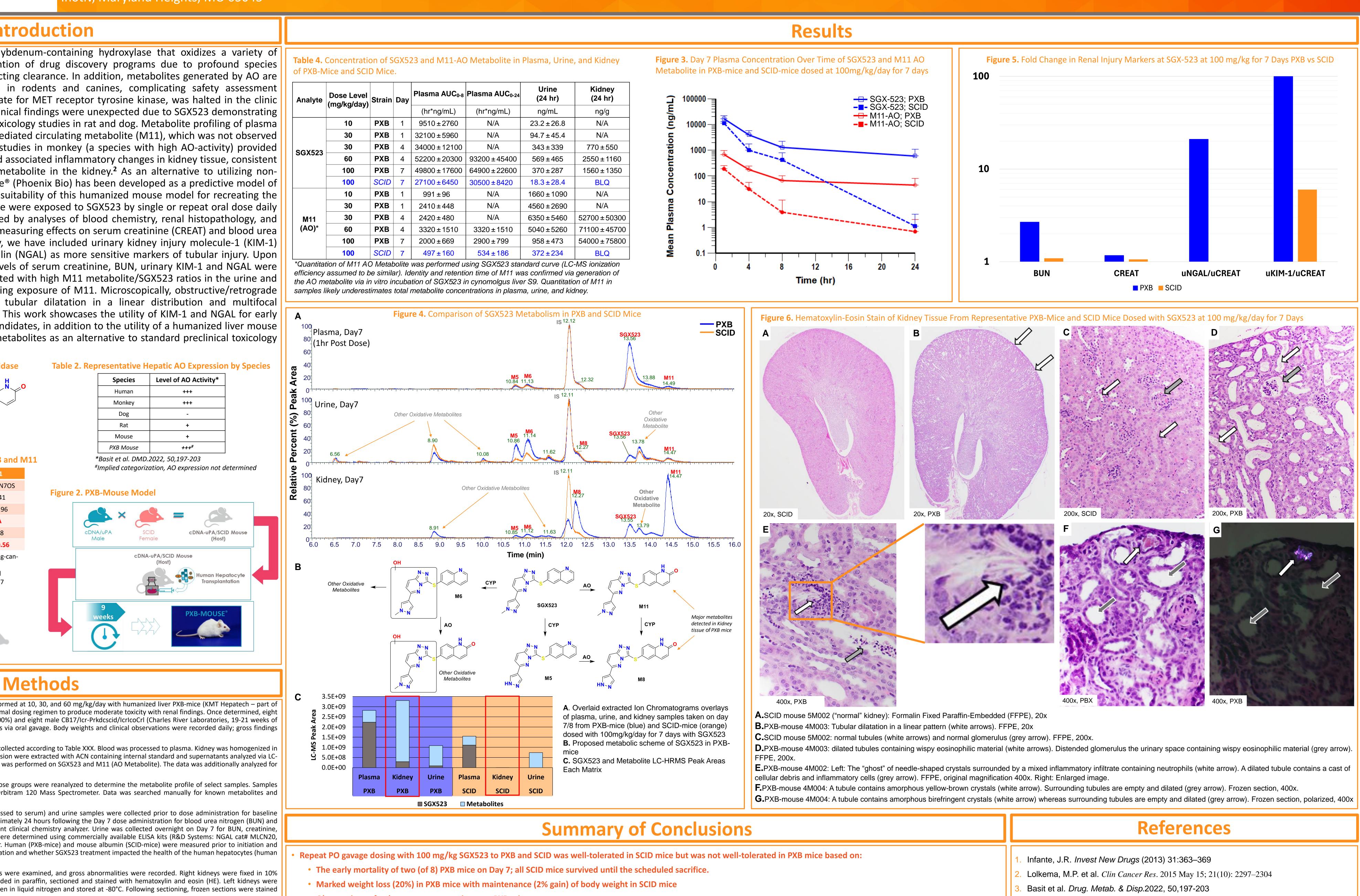


### Table 1. Predicted Phys Chem Properties of SGX523 and M11

Property	SGX523	M11						
Chemical Formula	C18H13N7S	C18H13N7OS						
MW	359.41	375.41						
LogP	3.0/2.74	2.7/1.96 <b>N/A</b> 93.78						
Quinoline pKa	4.2							
TPSA	73.81							
Solubility (µg/mL, in water)	2.5-5.4	0.13-0.56						
Refs: https://www.simulations-plus.com/resource/silicomodeling-can- predict-unforeseen-renal-failure-caused-sgx523-c-met-kinase- inhibitor/; https://www.molinspiration.com/cgi/properties ; and Diamond, S., et al, Drug Metab. And Disp., 2010, vol38 (8), p1277								



Species	Level of AO Ac			
Human	+++			
Monkey	+++			
Dog	-			
Rat	+			
Mouse	+			
PXB Mouse	+++#			
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### In Life Study Design: Dose range finding (DRF) studies were performed at 10, 30, and 60 mg/kg/day with humanized liver PXB-mice (KMT Hepatech – part of the PhoenixBio Group, 28-46 weeks of age) to determine the optimal dosing regimen to produce moderate toxicity with renal findings. Once determined, eight male PXB-mice (19-22 weeks of age, replacement indices = 82-100%) and eight male CB17/Icr-Prkdcscid/IcrIcoCrl (Charles River Laboratories, 19-21 weeks of age) were administered 100 mg/kg SGX-523 once daily for 7 days via oral gavage. Body weights and clinical observations were recorded daily; gross findings were documented at the time of sacrifice on Day 8.

Pharmacokinetic analysis: Blood, urine, and kidney tissues were collected according to Table XXX. Blood was processed to plasma. Kidney was homogenized in 70% IPA (1:3 tissue:solvent ratio by volume). The resulting suspension were extracted with ACN containing internal standard and supernatants analyzed via LC-HRMS with a Sciex 7600 Zeno-Tof Mass Spectrometer. Bioanalysis was performed on SGX523 and M11 (AO Metabolite). The data was additionally analyzed for other major metabolites (MS and MSMS).

**Metabolite Analysis:** Samples prepared for bioanalysis of high dose groups were reanalyzed to determine the metabolite profile of select samples. Samples were analyzed analyzed via LC-HRMS with a Thermo Exploris Orbitram 120 Mass Spectrometer. Data was searched manually for known metabolites and processed via MassMetasite software.

Clinical Pathology and Urinary Biomarkers: Whole blood (processed to serum) and urine samples were collected prior to dose administration for baseline data. Serum samples were collected at necropsy on Day 8 approximately 24 hours following the Day 7 dose administration for blood urea nitrogen (BUN) and creatinine analysis performed using a Randox Daytona instrument clinical chemistry analyzer. Urine was collected overnight on Day 7 for BUN, creatinine, NGAL, and KIM-1 analysis. Concentrations of KIM-1 and NGAL were determined using commercially available ELISA kits (R&D Systems: NGAL cat# MLCN20, KIM-1 cat# MKM100) analyzed on a SpectraMax384 plate reader. Human (PXB-mice) and mouse albumin (SCID-mice) were measured prior to initiation and following completion of dosing to monitor the degree of humanization and whether SGX523 treatment impacted the health of the human hepatocytes (human albumin was stable during study, data not shown here).

Necropsy and Pathology At a limited necropsy, liver and kidneys were examined, and gross abnormalities were recorded. Right kidneys were fixed in 10% neutral buffered formalin, trimmed, processed routinely, embedded in paraffin, sectioned and stained with hematoxylin and eosin (HE). Left kidneys were embedded in optimal cutting temperature (OCT compound), frozen in liquid nitrogen and stored at -80°C. Following sectioning, frozen sections were stained with HE. Liver observations not reported here.

### Table 3. In Life Study Design

Study Phase Animals		Dose Level (mg/kg) Days		Sample	Time Collections (Hours Postdose)		
	Animais		Collection	Plasma	Urine	Kidney	
1	PXB-Mice	10	1	Day 1	0.5, 1, 2, 4, 8	24	N/A
2	PXB-Mice	30	4	Day 1 & 4	0.5, 2, 8	24	Terminal
3	PXB-Mice	60	4	Day 4	0.5, 2, 8, 24	24	Terminal
4	PXB-Mice	100	7	Day 7	1, 4, 8, 24	24	Terminal
	SCID-Mice	100	7	Day 7	1, 4, 8, 24	24	Terminal

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• Observations of unkempt appearance in some PXB mice • The level of AO metabolism in the PXB-mouse was more representative of human metabolism than the control (SCID) mice Observed dose proportional increases in plasma and renal exposure of SGX523 and M11 (and other metabolites) in the PXB-mice In PXB mice, microscopic findings of tubular dilatation, casts, inflammatory infiltrates and tubular basophilia (not shown) were observed and correlated to increases in serum BUN CREAT, and urinary NGAL and KIM-1. Intralesional crystals in sections examined were consistent with high renal concentrations of M11.

These observations were collectively indicative of renal injury and impaired renal function in PXB mice dosed with SGX-523, not observed in similarly dosed SCID-mice. Comparison of in vivo metabolites reported with in vitro incubations using liver homogenates of collected tissues from unused PXB-mice and SCID-mice planned (not dosed).

Diamond, S., et al, *Drug Metab. And Disp.*, 2010, vol38 (8), p1277

Physcial chemical properties calculated from:https://www.simulations-

plus.com/resource/silicomodeling-can-predict-unforeseen-renal-failure-caused-sgx523-cmet-kinase-inhibitor/ and https://www.molinspiration.com/cgi/properties

# Acknowledgements

We would like to thank PhoenixBio for providing the PXB-mice used in the reported studies.